



Biogeochemical Controls Over Organohalide-Respiring Chloroflexi



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Contaminated Sites in the U.S.



- 1,322 Superfund sites ●
- 3,747 RCRA sites ●
- >450,000 Brownfields ●



166 MILLION
PEOPLE

Live
Within

3
MILES

of a Superfund or a
RCRA Corrective
Action Site, Equal to

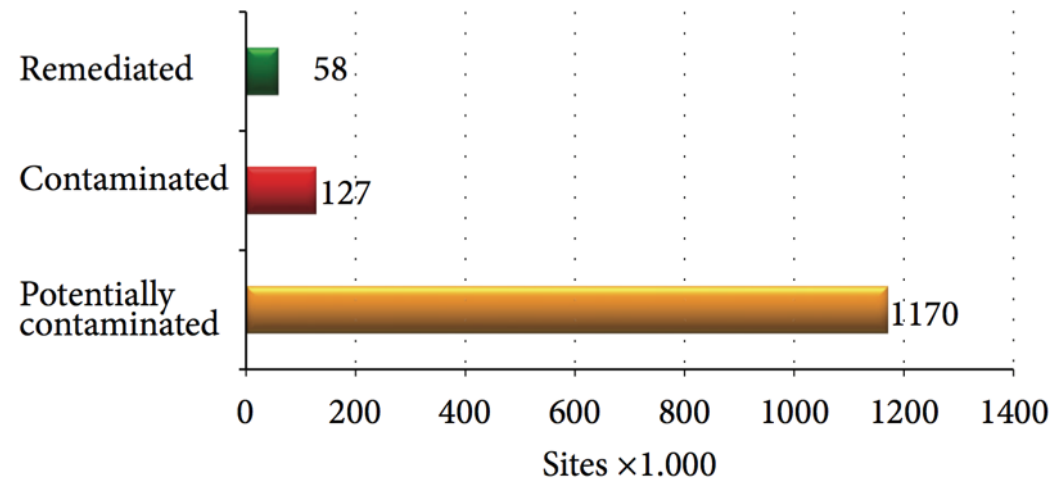
53%
OF THE U.S.
POPULATION

Including

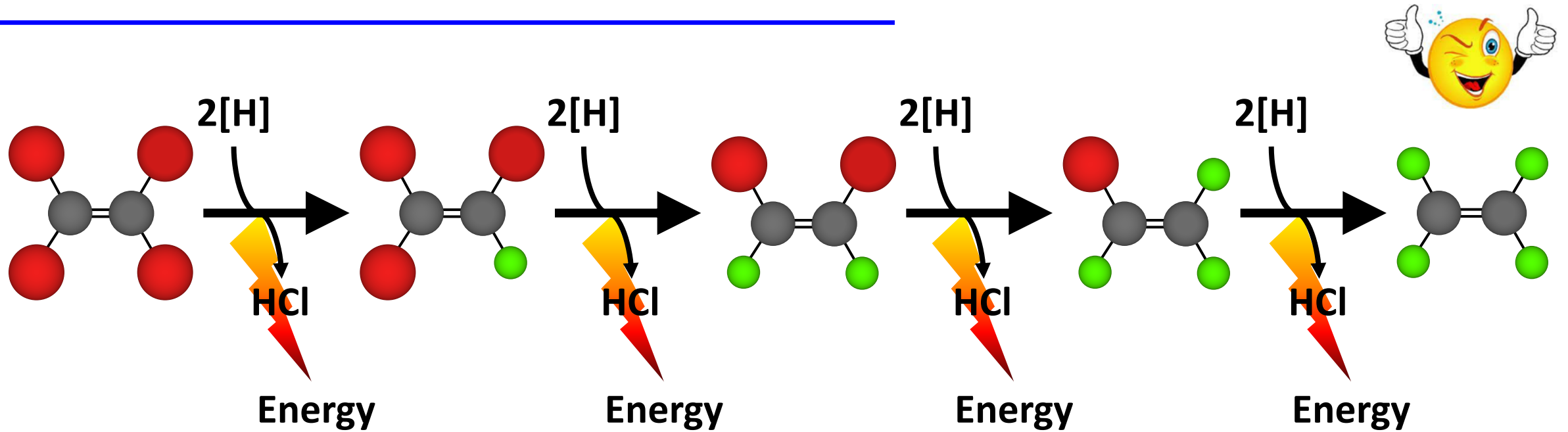
55% OF ALL
CHILDREN
UNDER AGE 5

... and in Europe

Majority of sites impacted
with chlorinated compounds



Reductive Dechlorination: A Process that Leads to Contaminant Detoxification *In Situ*

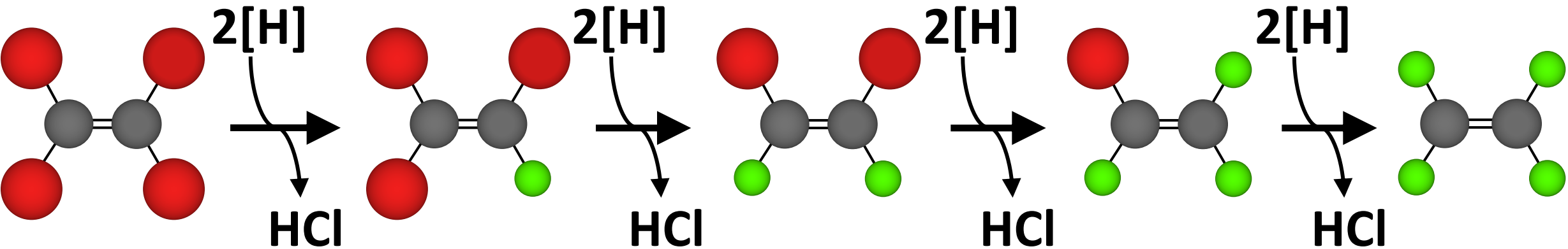


Organohalide Respiration

Freedman, D. L., and J. M. Gossett. 1989. *Appl. Environ. Microbiol.* 55:2144-2151
He et al. 2003. *Nature.* 424:62-65

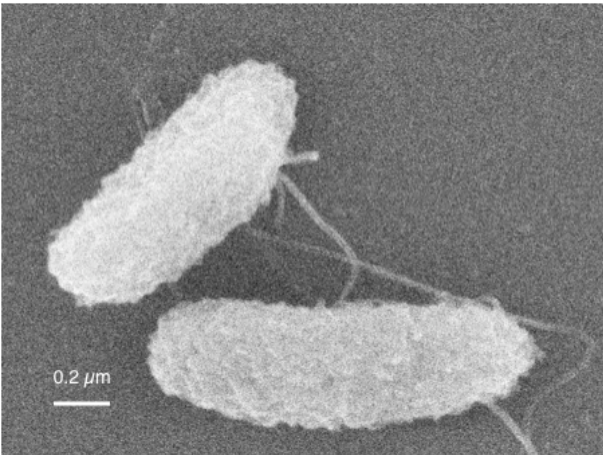


Populations Involved in Reductive Dechlorination of CEs

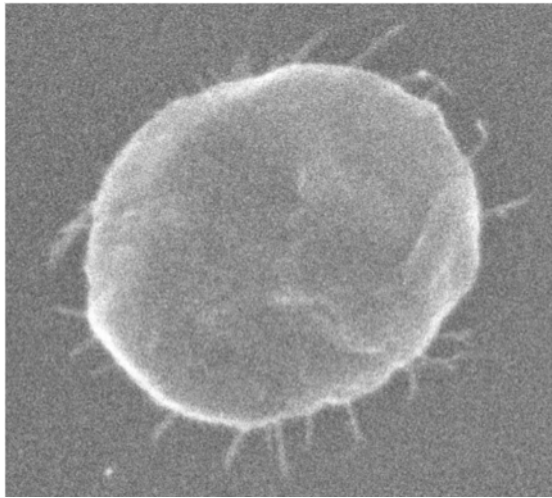


Geobacter lovleyi, Dehalobacter, Sulfurospirillum, Desulfuromonas, Desulfitobacterium

Dehalococcoides mccartyi



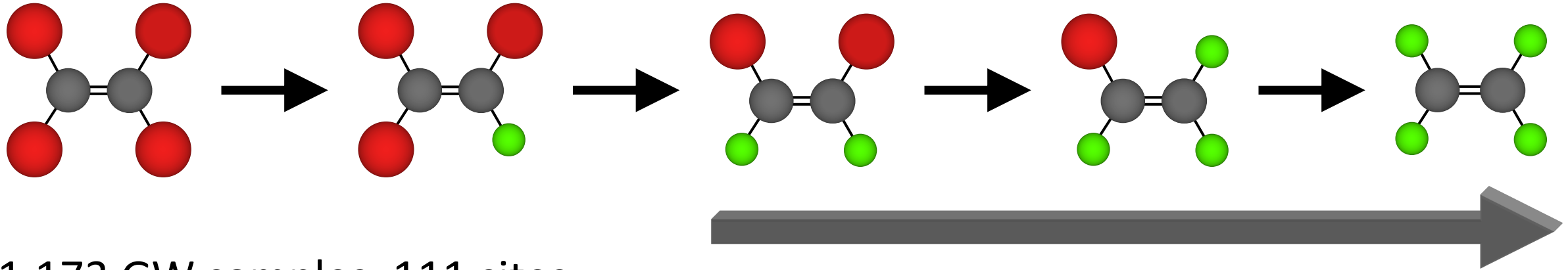
Sung et al. 2006
AEM, 72:2775



Löffler et al. 2013
IJSEM, 63:625



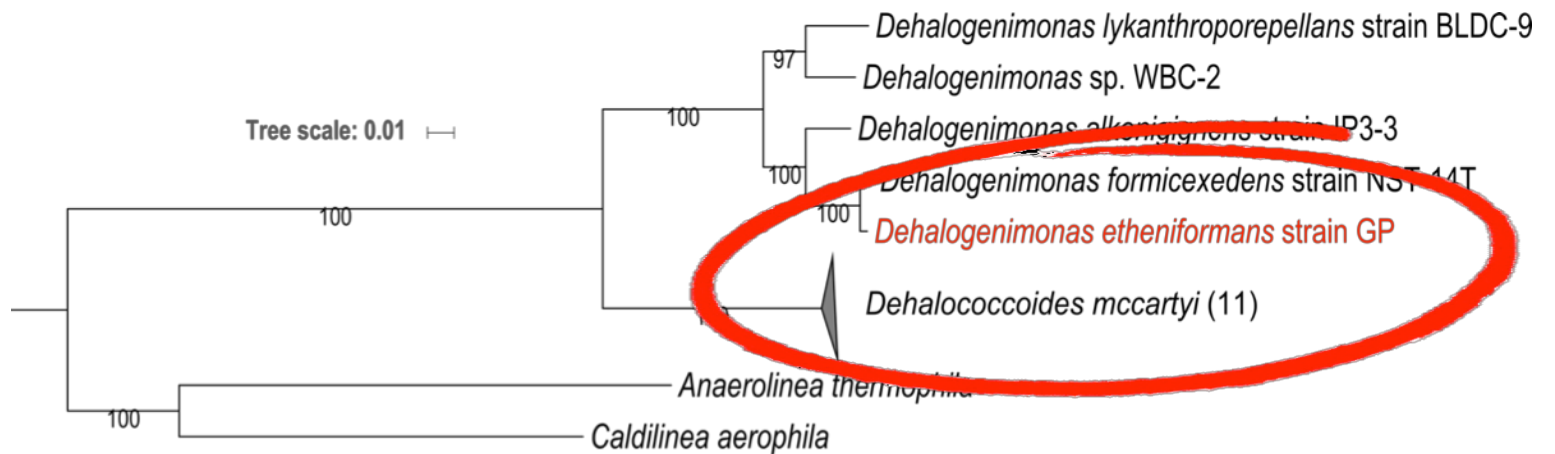
Populations Involved in Reductive Dechlorination of Chlorinated Ethenes



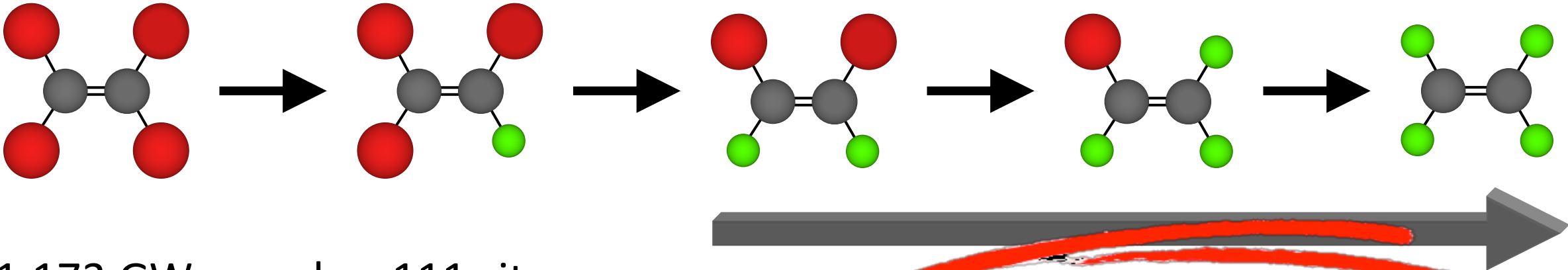
1,173 GW samples, 111 sites

849 samples: *Dhc* & *Dhgm*

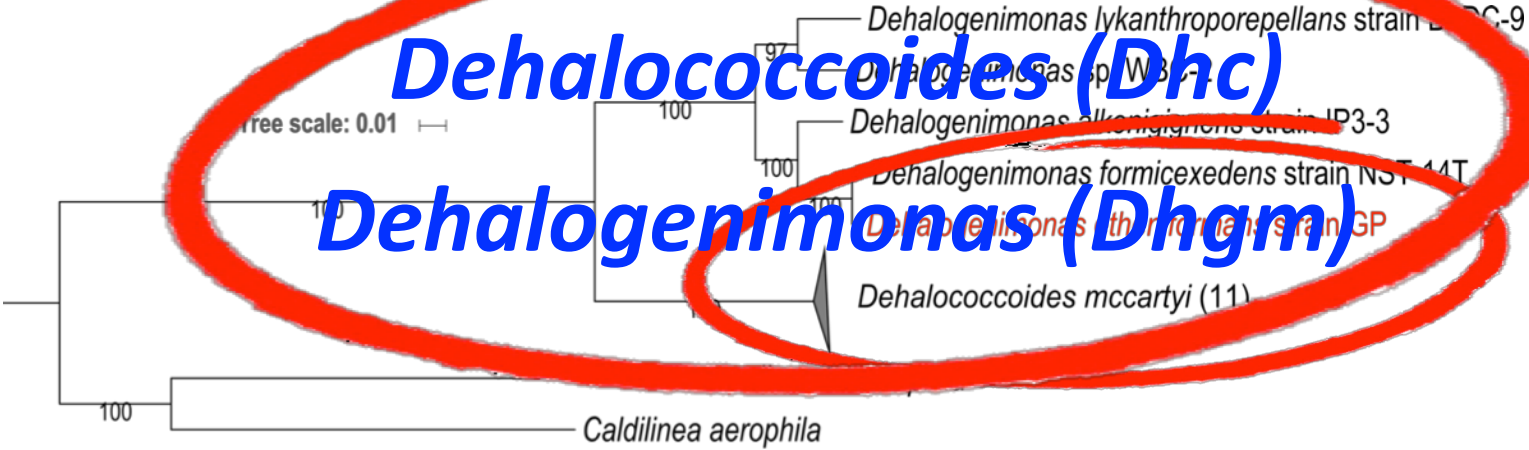
65%: *Dhgm* outnumber *Dhc*



Populations Involved in Reductive Dechlorination of Chlorinated Ethenes



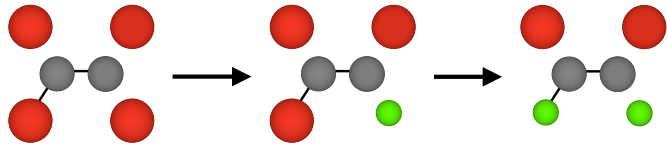
1,173 GW samples, 111 sites
 849 samples: *Dhc* & *Dhgm*
 65%: *Dhgm* outnumber *Dhc*



Structural Basis of Organohalide Respiration

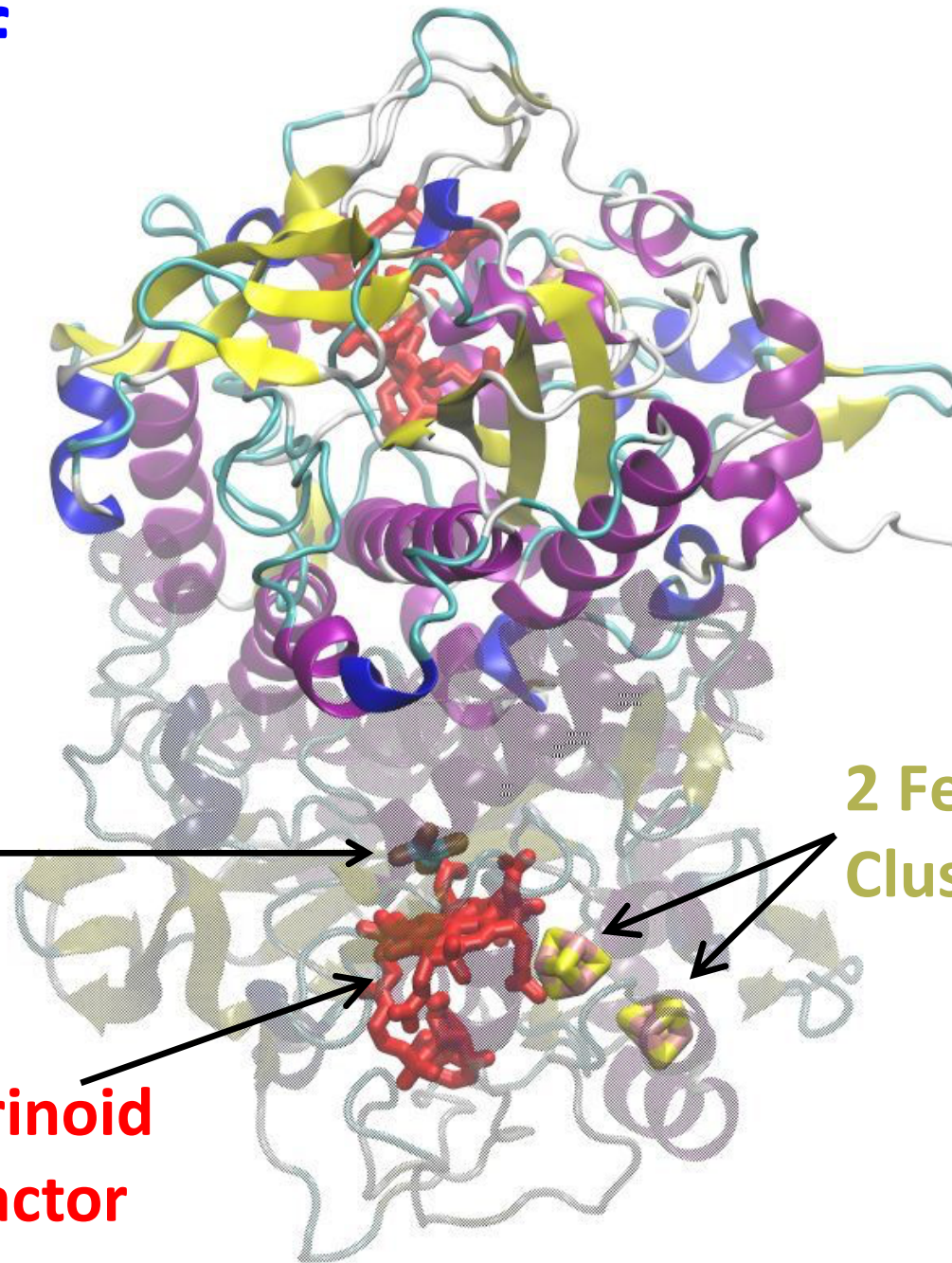
Dimer of PceA

Sulfurospirillum multivorans



Bommer et al. 2014. Science, 346:455

Payne et al. 2015. Nature, 517:513



Corrinoid Cofactor

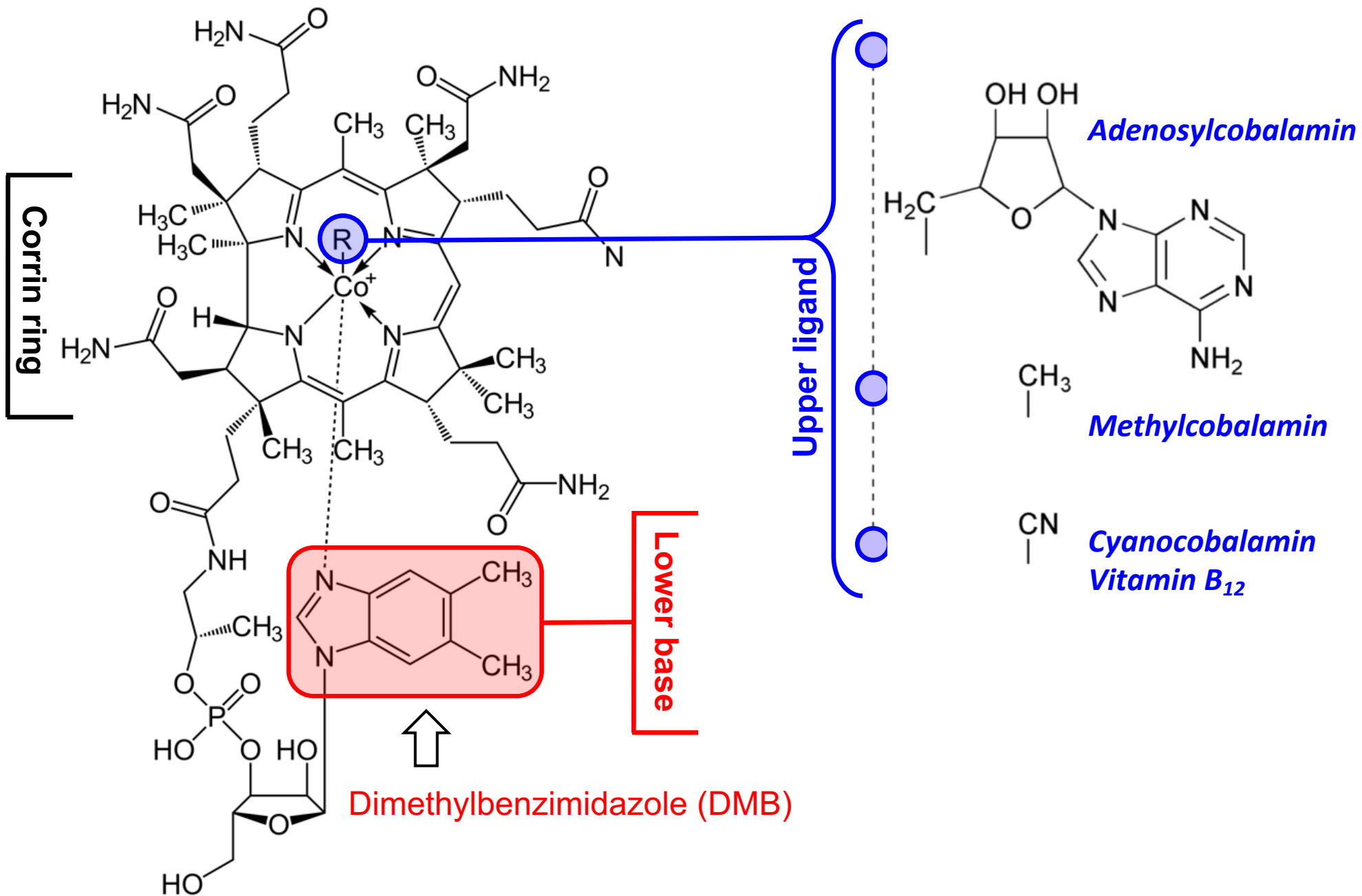
2 FeS Clusters

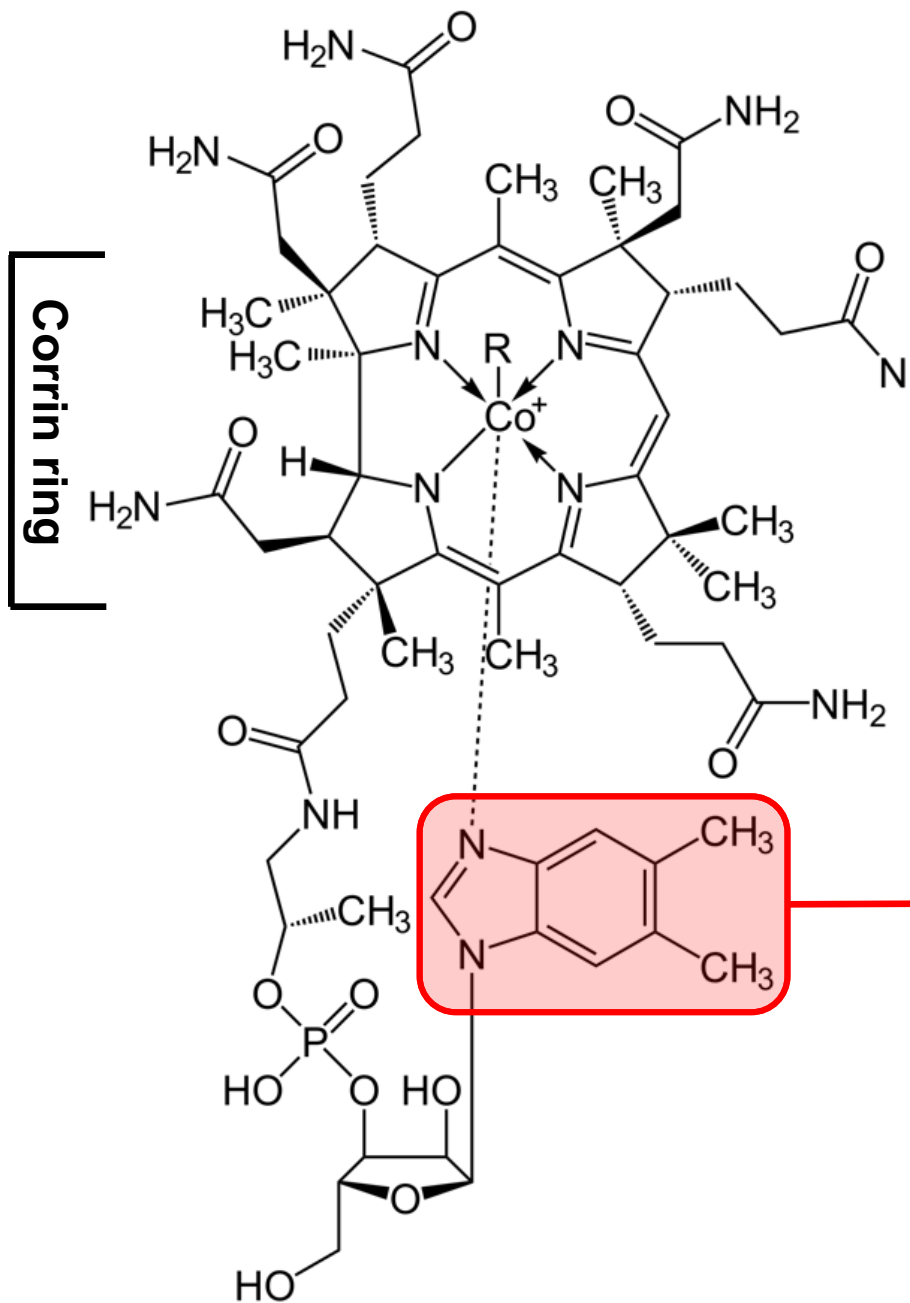
Lorenz Adrian · Frank E. Löffler *Editors*

Organohalide-Respiring Bacteria

Springer

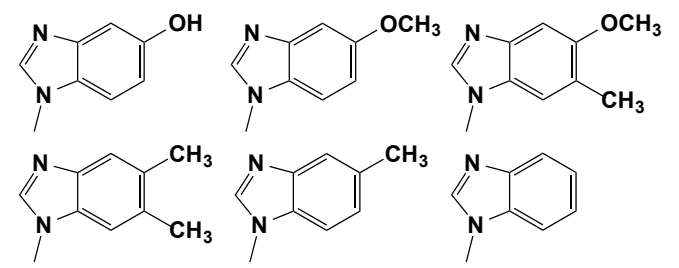




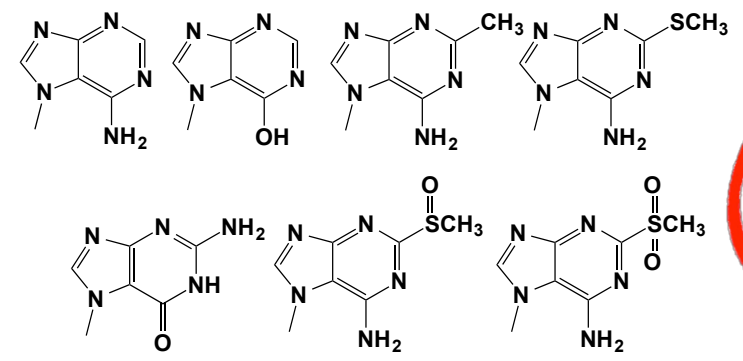


Lower Bases

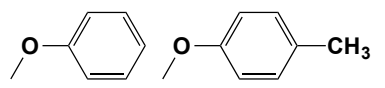
Benzimidazole (Bza) type



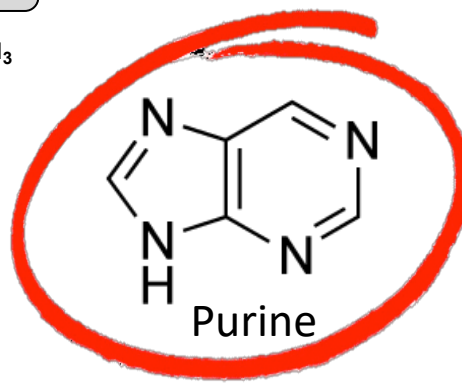
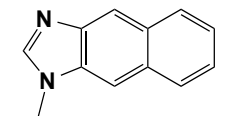
Nucleobase type



Phenol type



Naphthimidazole

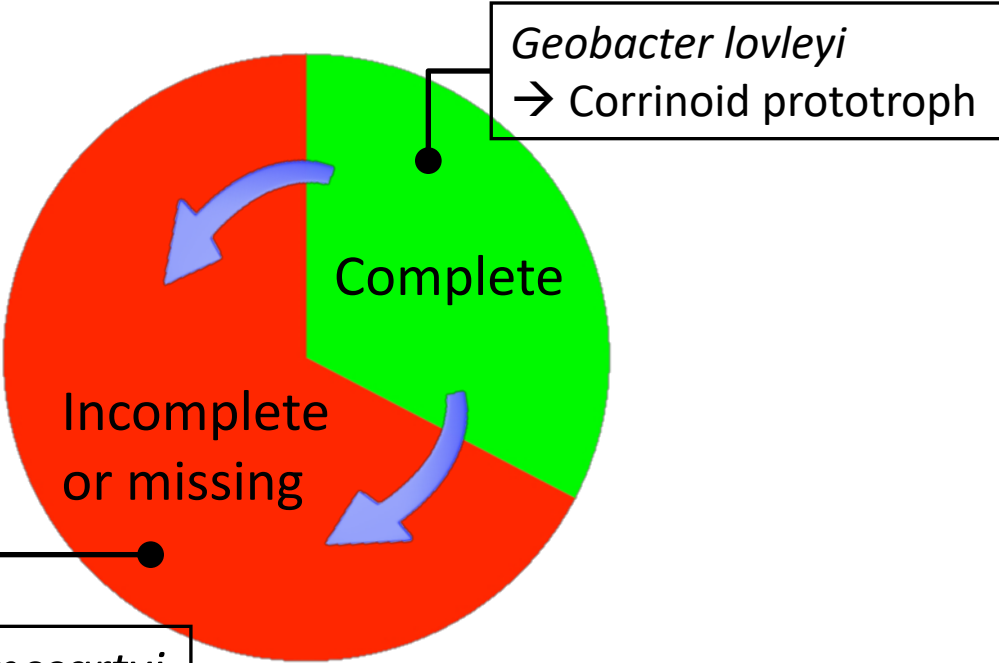


Yan et al. 2018.
Nat. Chem. Biol.
14:8-14.

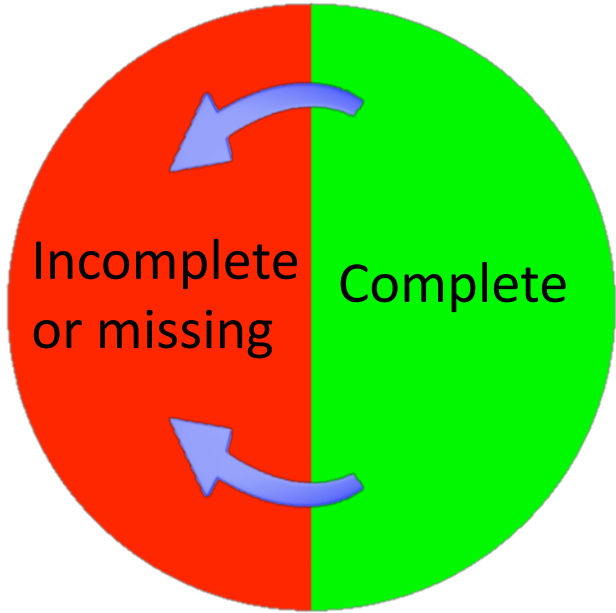


De novo Biosynthesis of Corrinoids

Bacteria
(n = 56,902)



Archaea
(n = 1,362)



Dehalococcoides mccartyi
→ Corrinoid auxotroph

Geobacter lovleyi
→ Corrinoid prototroph



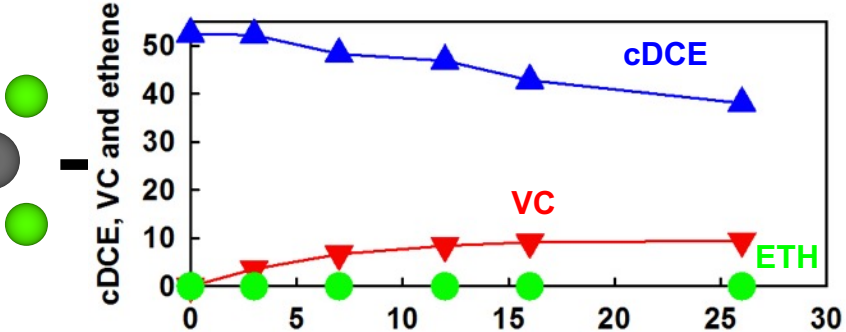
Dhc & Dhgm: Strict Requirement for Corrinoid

Dhc strain BAV1

No B₁₂

cDCE

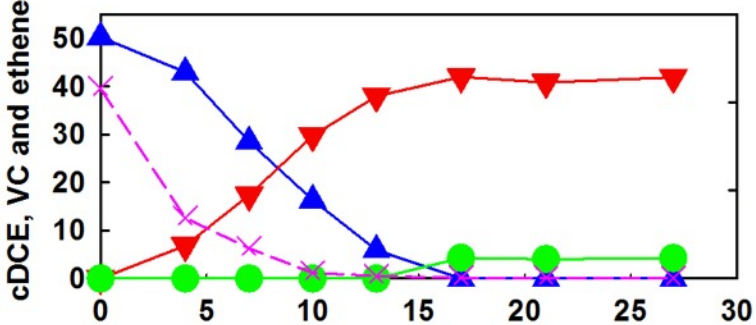
VC



No dechlorination



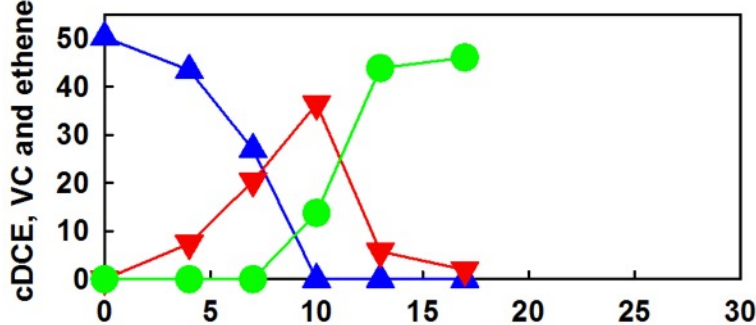
Limited B₁₂ [1 µg/L]



VC stall



Sufficient B₁₂ [25 µg/L]



Complete Dechlorination (Detoxification)

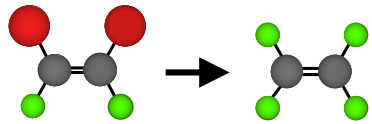


Specific Aims

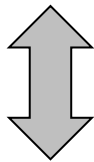
- **Aim 1:** Explore the specific cobamide requirements of organohalide-respiring *Dhc* relevant for detoxification of chlorinated ethenes
- **Aim 2:** Demonstrate that geochemical conditions affect the specific cobamide pool, and hence *Dhc* activity
- **Aim 3:** Identify community and *Dhc* biomarkers that indicate when cobamide and/or lower base bioavailability limit *Dhc* reductive dechlorination activity



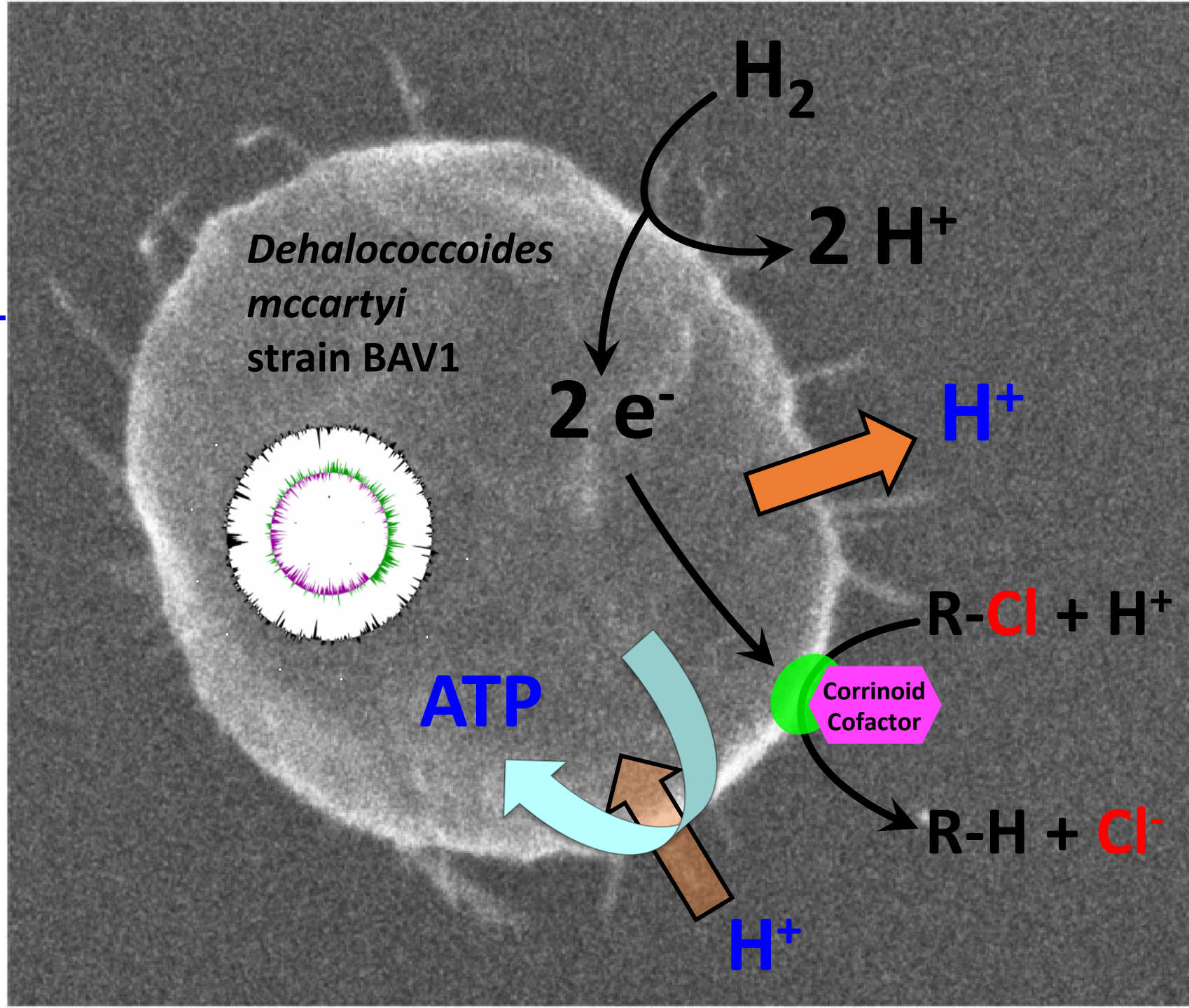
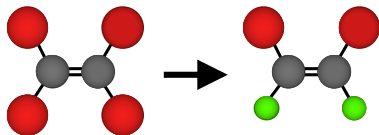
Simplified Model of Organohalide Respiration



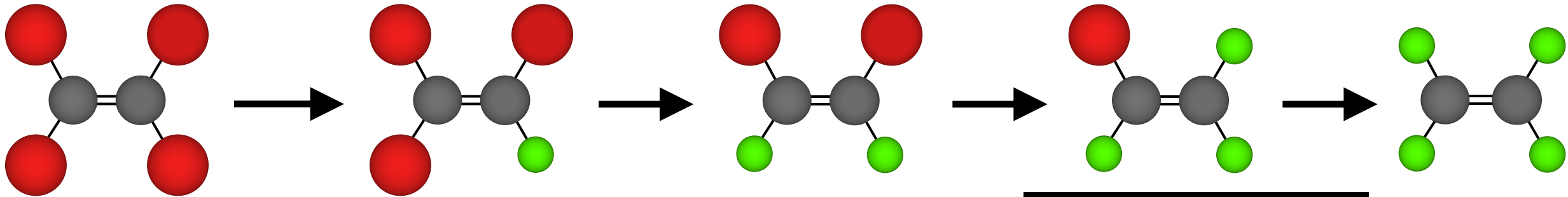
Dehalococcoides mccartyi
→ Corrinoid auxotroph



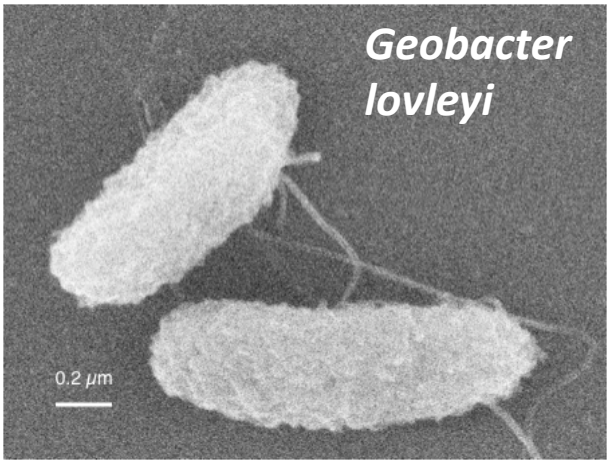
Geobacter lovleyi
→ Corrinoid prototroph



Who Supplies Corrinoid to *Dhc* and *Dhgm*?

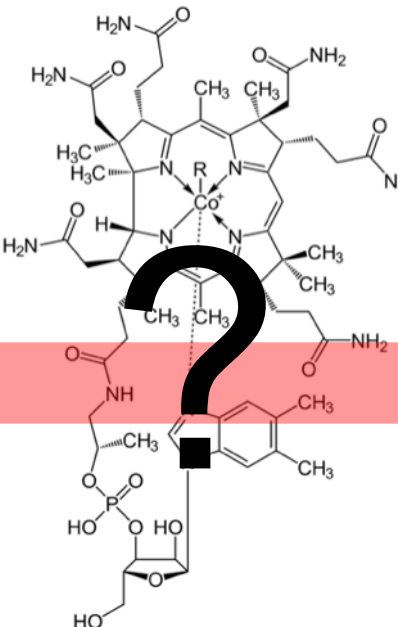


Dehalococcoides mccartyi



Geobacter lovleyi

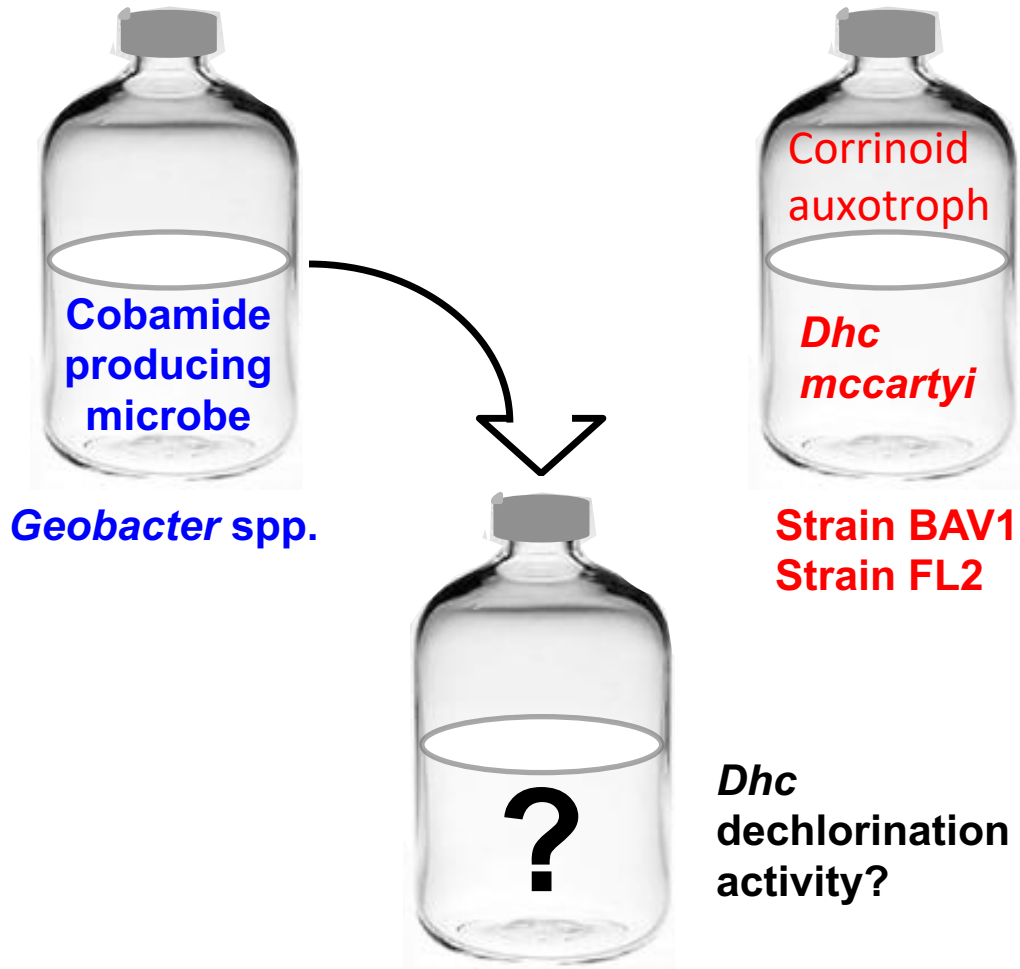
Corrinoid +



Corrinoid -



Co-Culture Experiments: Corrinoid Producer / *Dhc mccartyi*



Summary of Co-Culture Experiments

Co-Cultures		<i>Dhc</i> Growth
	<i>Dhc</i> Strains	
	BAV1, FL2	+



Geobacter spp.



Strain BAV1
Strain FL2

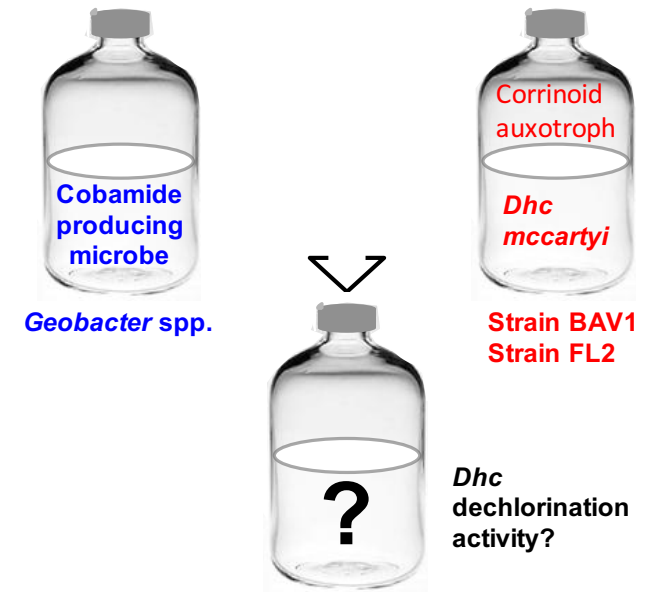


Dhc dechlorination activity?



Summary of Co-Culture Experiments

Co-Cultures		<i>Dhc</i> Growth
Corrinoid Producer	<i>Dhc</i> Strains	
	BAV1, FL2	+
	BAV1, FL2	-
	BAV1, FL2, GT	-
	BAV1, FL2	-
	BAV1, FL2	-
	BAV1, FL2, GT	-



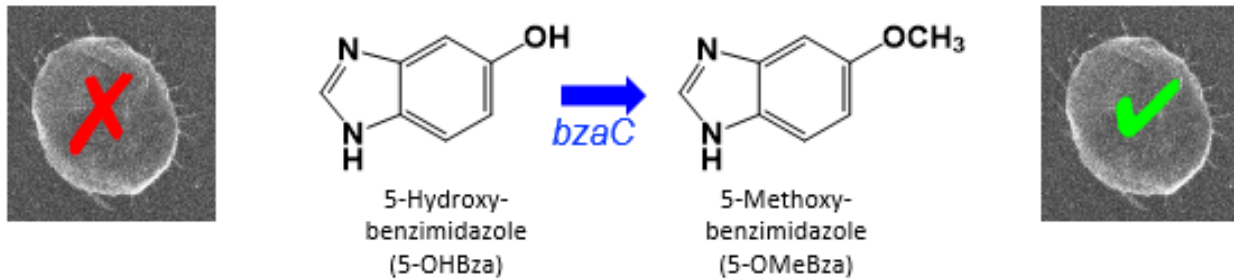
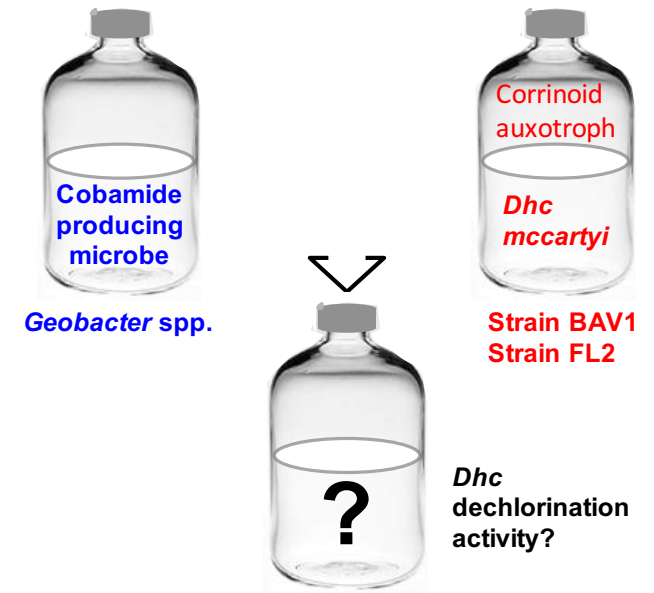
Yan et al. 2012. Appl. Environ. Microbiol. 78:6630-6636

Yan et al. 2013. Phil. Trans. R. Soc. B. 368, 20120320



Co-Culture Experiments

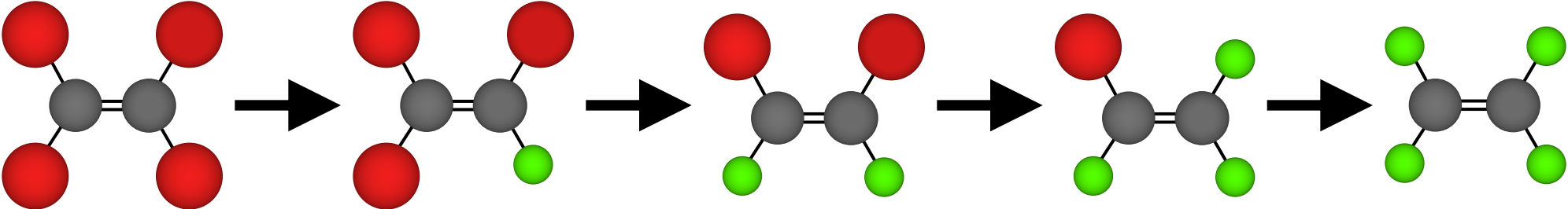
Co-Cultures		<i>Dhc</i> Growth
Corrinoid Producer	<i>Dhc</i> Strains	
<i>Geobacter lovleyi</i>	BAV1, FL2	+
<i>Geobacter sulfurreducens</i> Wildtype	BAV1, FL2	-
<i>Geobacter sulfurreducens</i> + pNJ052	BAV1, FL2	+



Jiang et al. 2019. In Preparation



Key *Dhc* RDases



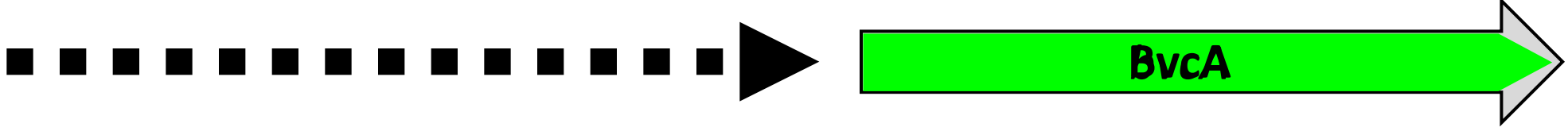
Dhc str. 195



Dhc str. FL2



Dhc str. BAV1



Dhc str. VS

Dhc str. GT

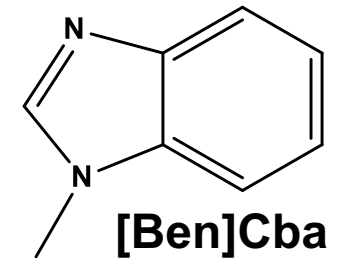
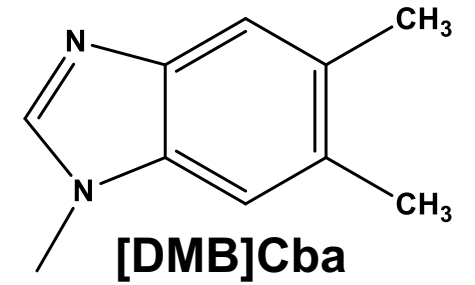
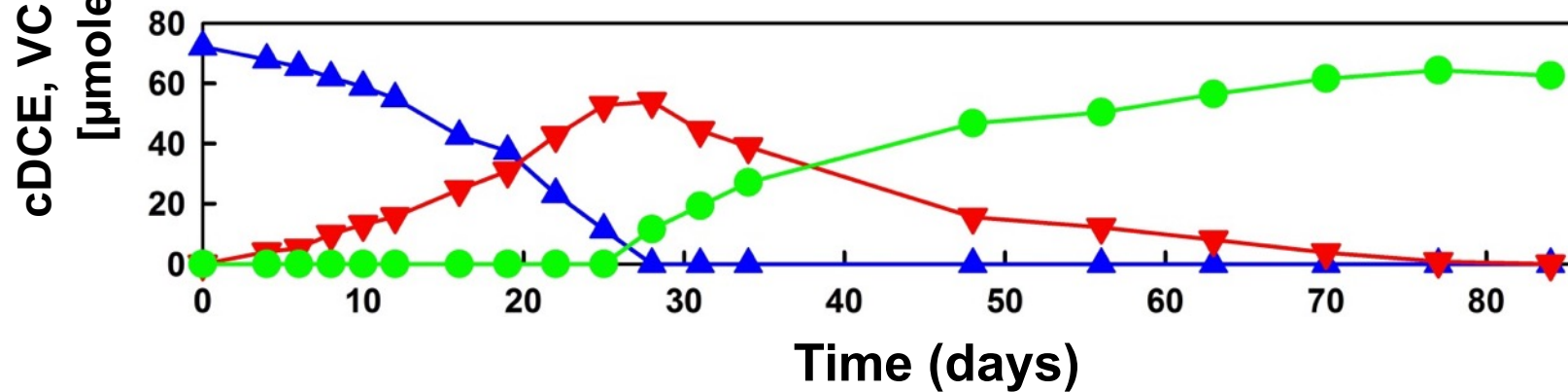
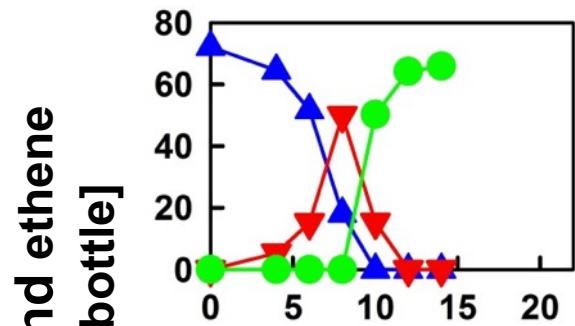


Lower Base Affects Dechlorination Activity

Strain BAV1 (BvcA)

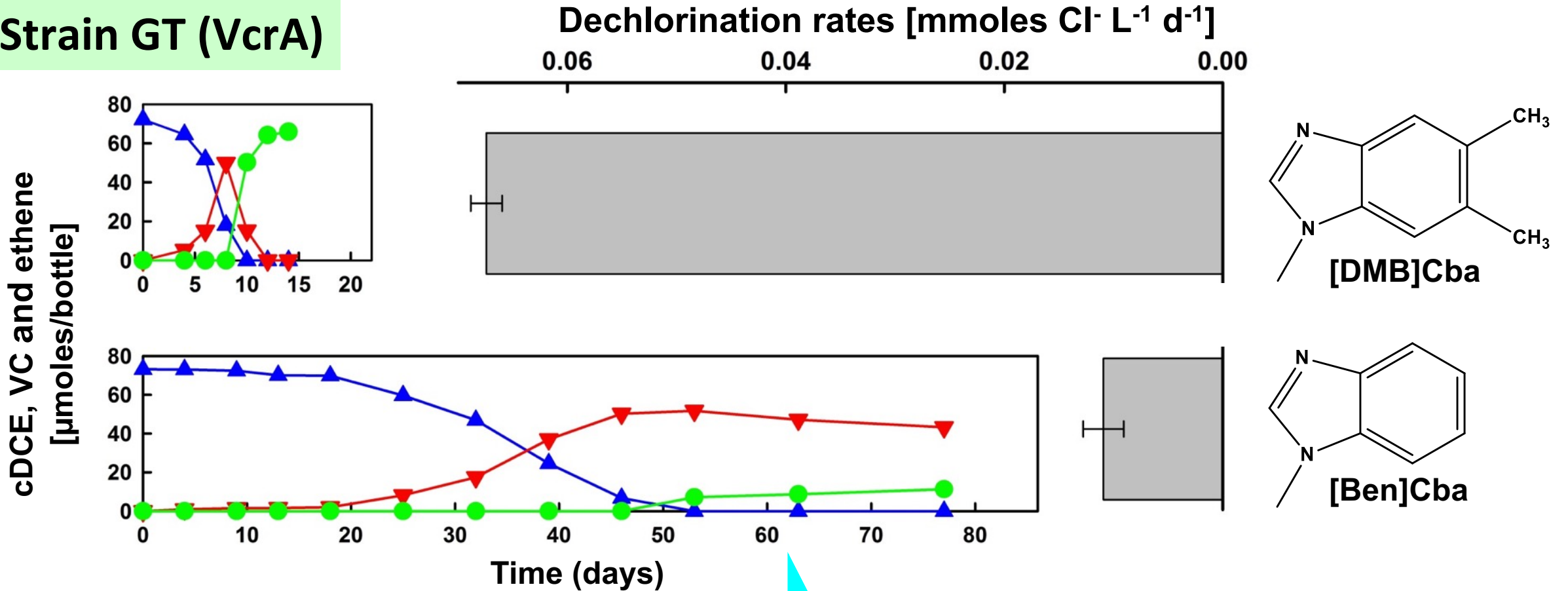
Dechlorination rates [$\text{mmoles Cl}^- \text{L}^{-1} \text{d}^{-1}$]

0.12 0.10 0.08 0.06 0.04 0.02 0.00



Lower Base Affects Dechlorination Activity

Strain GT (VcrA)



Yan et al. 2016. ISME J. 10:1092–1101



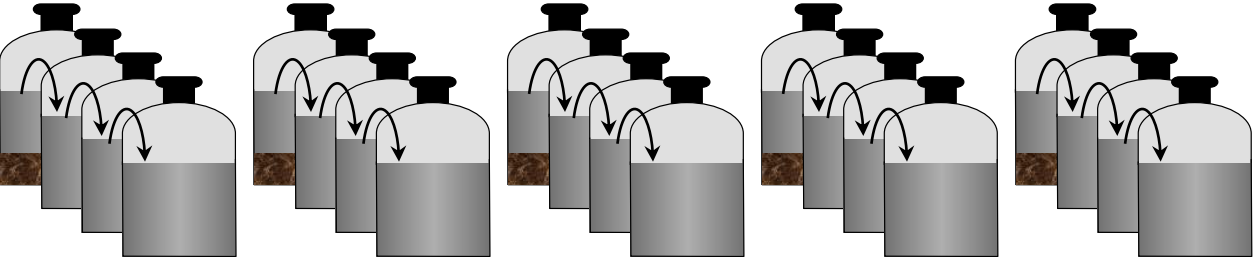
Lower base affects dechlorination rates and endpoints



Corrinoid Production Under Different Redox Conditions

Third Creek Site
Knoxville, TN

- Metal-manufacturing
- Chlorinated solvents



Fermenting
Nitrate-Reducing
Iron-Reducing
Sulfate-Reducing
Methanogenic

Commerce Street Superfund Site
Williston, Vermont

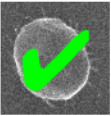
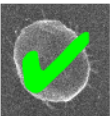
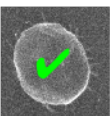
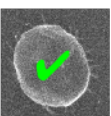
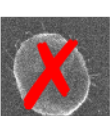
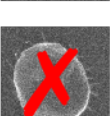
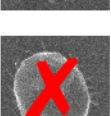
- Multi-tenant industrial park
- TCE, *cis*-DCE, petroleum hydrocarbons, metals (chromium, cadmium, nickel)



Corrinoid Extraction and Identification

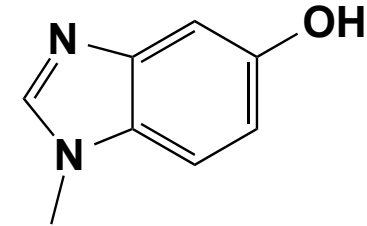
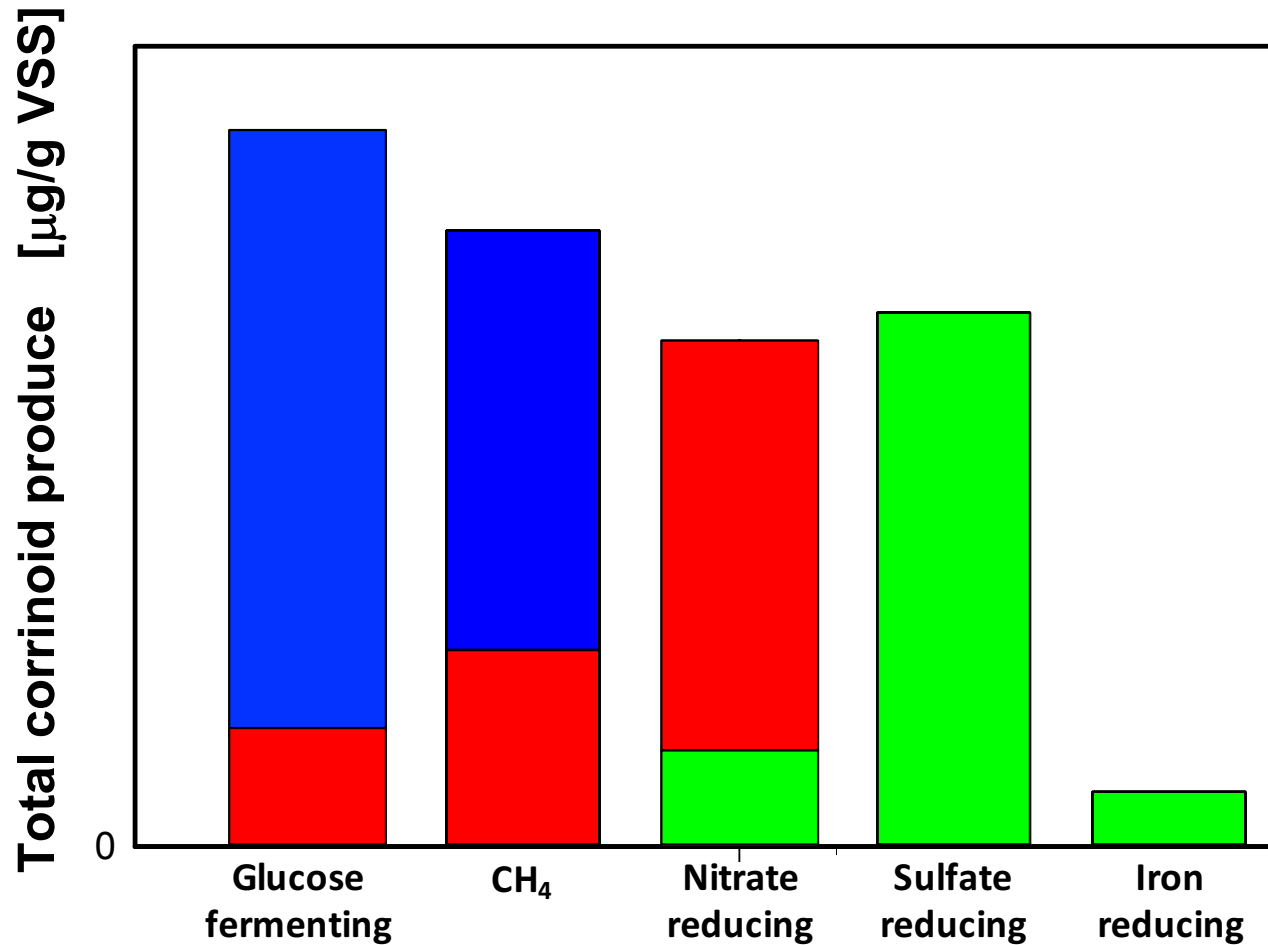


Corrinoids Produced by the Community Under Different Redox Conditions

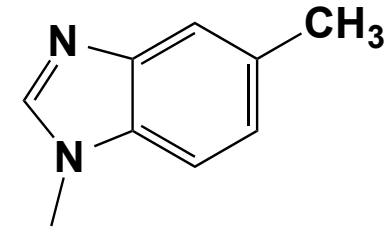
	Corrinoids produced	Glucose fermenting	Methanogenic	Iron reducing	Sulfate reducing	Nitrate reducing
	<chem>Cc1c(C)c2c(c1)n[nH]2</chem>	-	-	+	+	+
	<chem>Cc1ccc2c(c1)n[nH]2</chem>	+	+	-	-	-
	<chem>COC1=CC=CC=C1n1c[nH]1</chem>	-	-	-	-	+
	<chem>C1=CC=C2C(=C1)N=CN2</chem>	-	-	-	-	-
	<chem>Oc1ccc2c(c1)n[nH]2</chem>	+	+	-	-	-
	<chem>COc1ccc(C)cc1</chem>	-	+	-	-	-
	<chem>COc1ccccc1</chem>	-	+	-	-	-



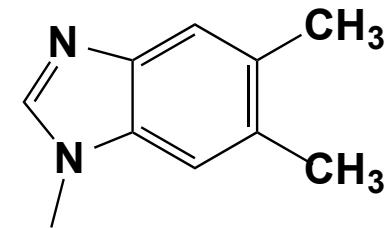
Redox Conditions Affect Corrinoid Type(s) and Quantity



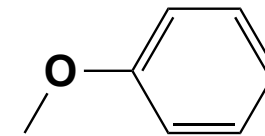
5-OHBza-Cba



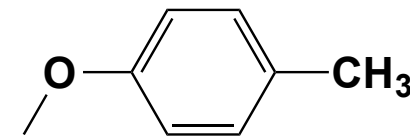
MeBza-Cba



DMB-Cba



Phe-Cba

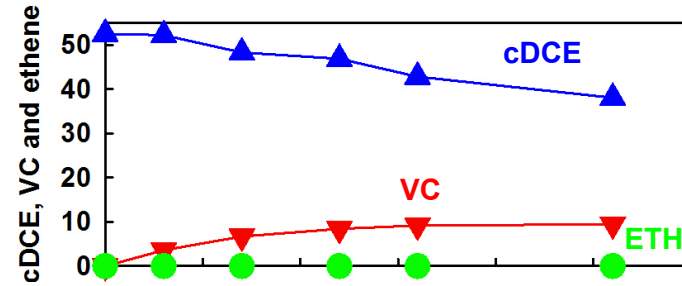


Cre-Cba

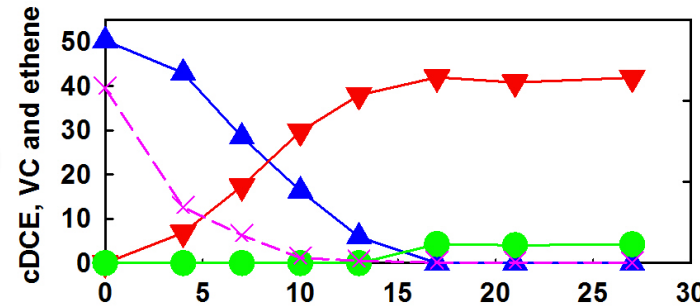


Corrinoid Quantity and Quality Determine *Dhc* Activity

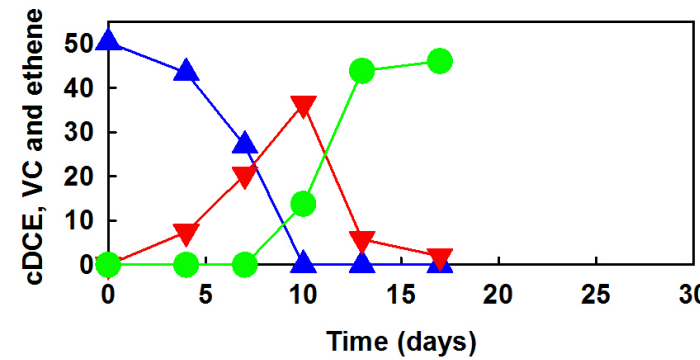
No B₁₂



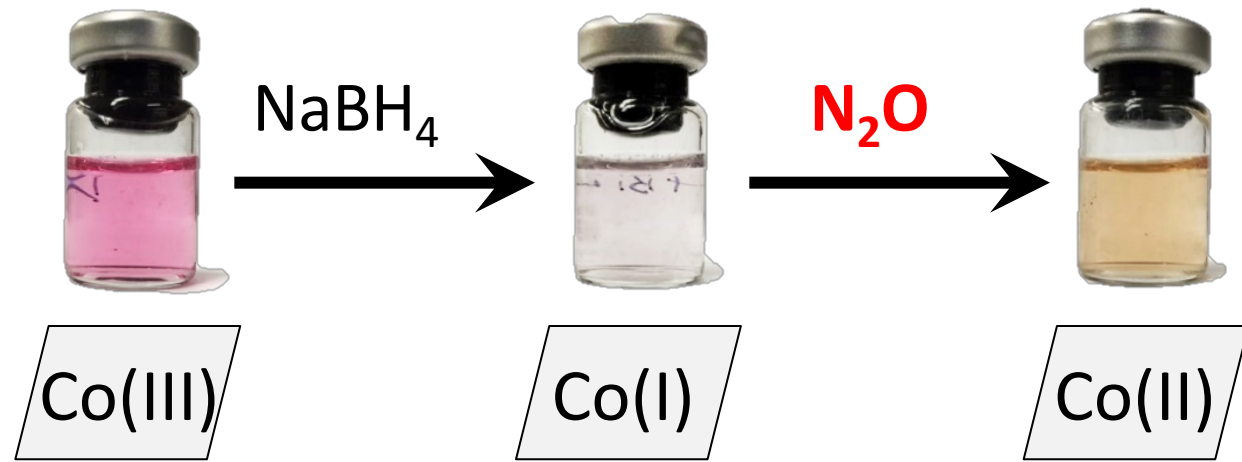
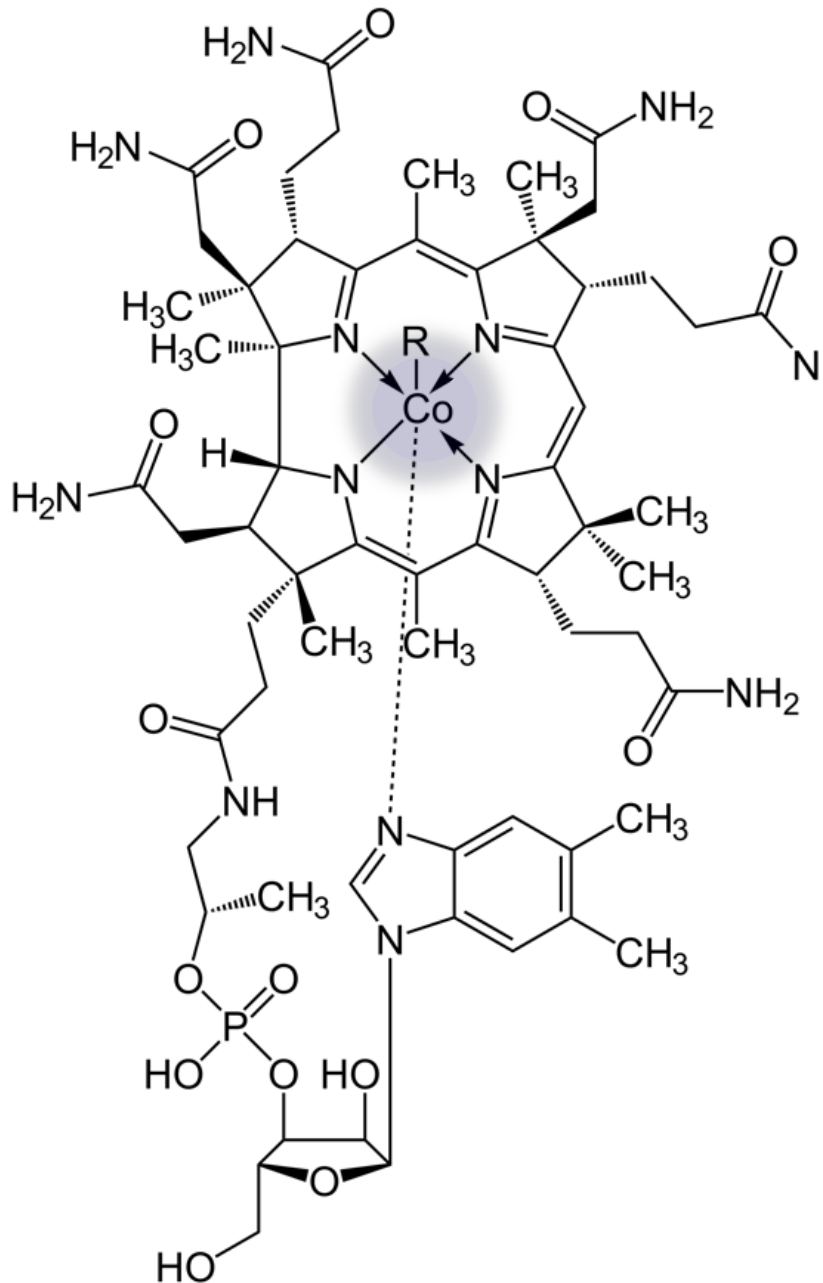
Limited B₁₂
[1 µg/L]



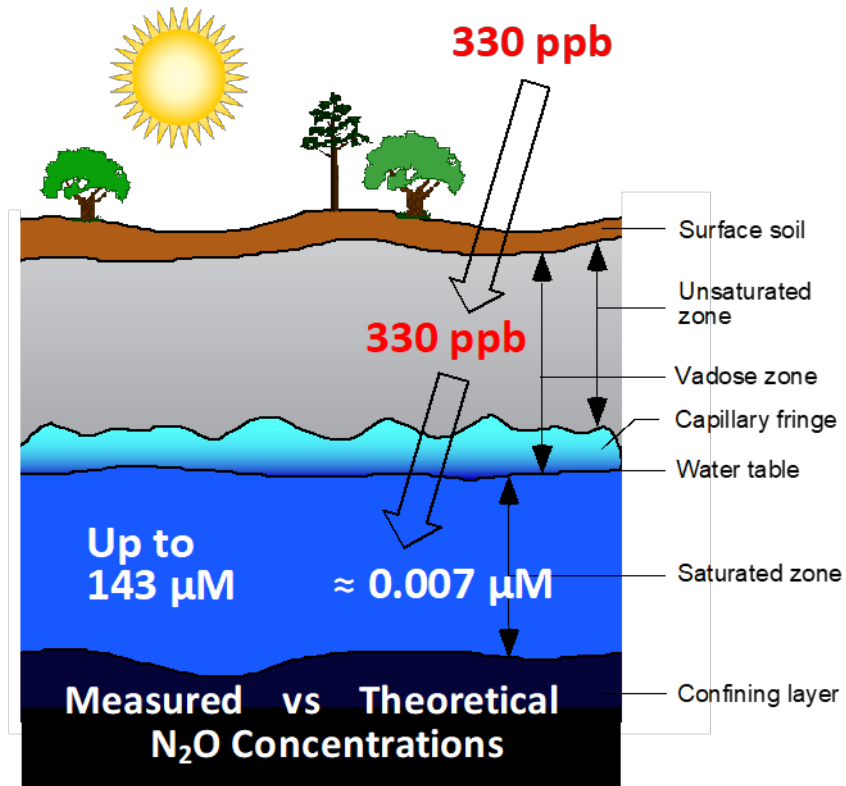
Sufficient B₁₂
[25 µg/L]



	<chem>Cc1c(C)nc2c1n[nH]2</chem>
	<chem>Cc1ccc2c(c1)n[nH]2</chem>
	<chem>COC1=CC=C2C(=C1)N=CN2</chem>
	<chem>CN1C=NC2=CC=CC=C12</chem>
	<chem>Oc1ccc2c(c1)n[nH]2</chem>
	<chem>Cc1ccc(OC)cc1</chem>
	<chem>COc1ccccc1</chem>



Banks et al. 1968. J. Chem. Soc. A, 2886

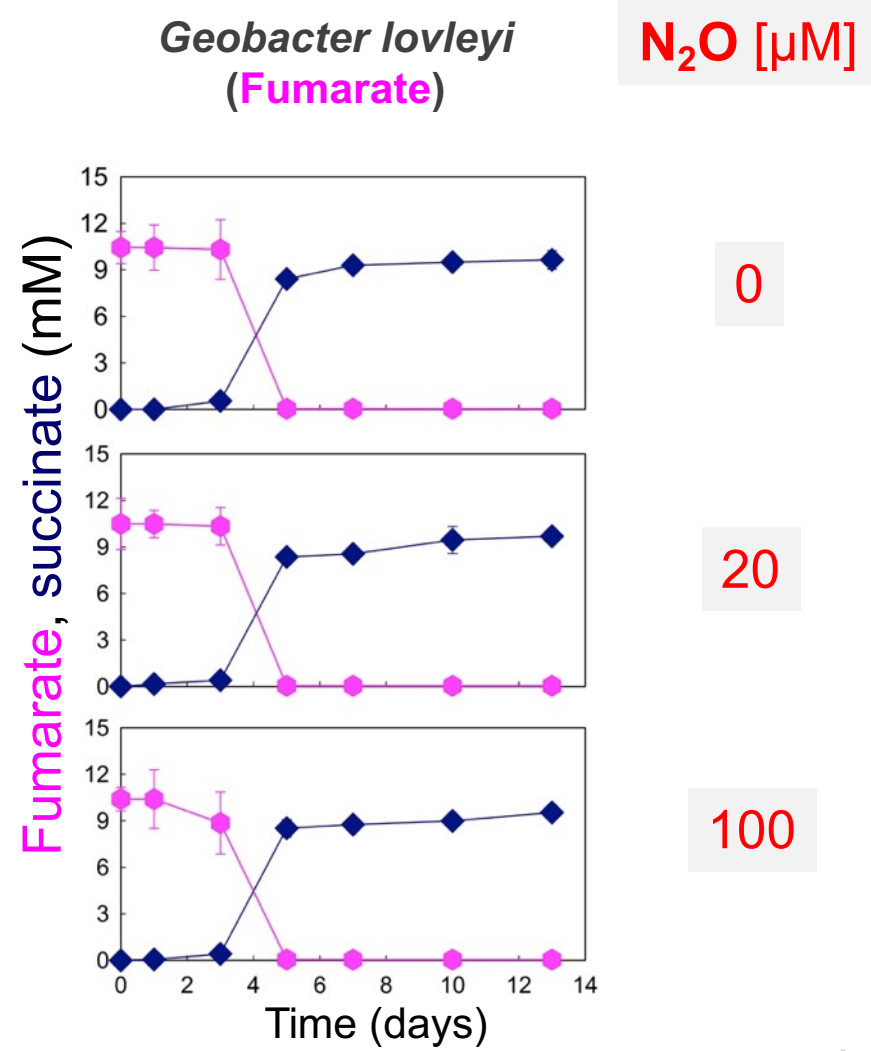
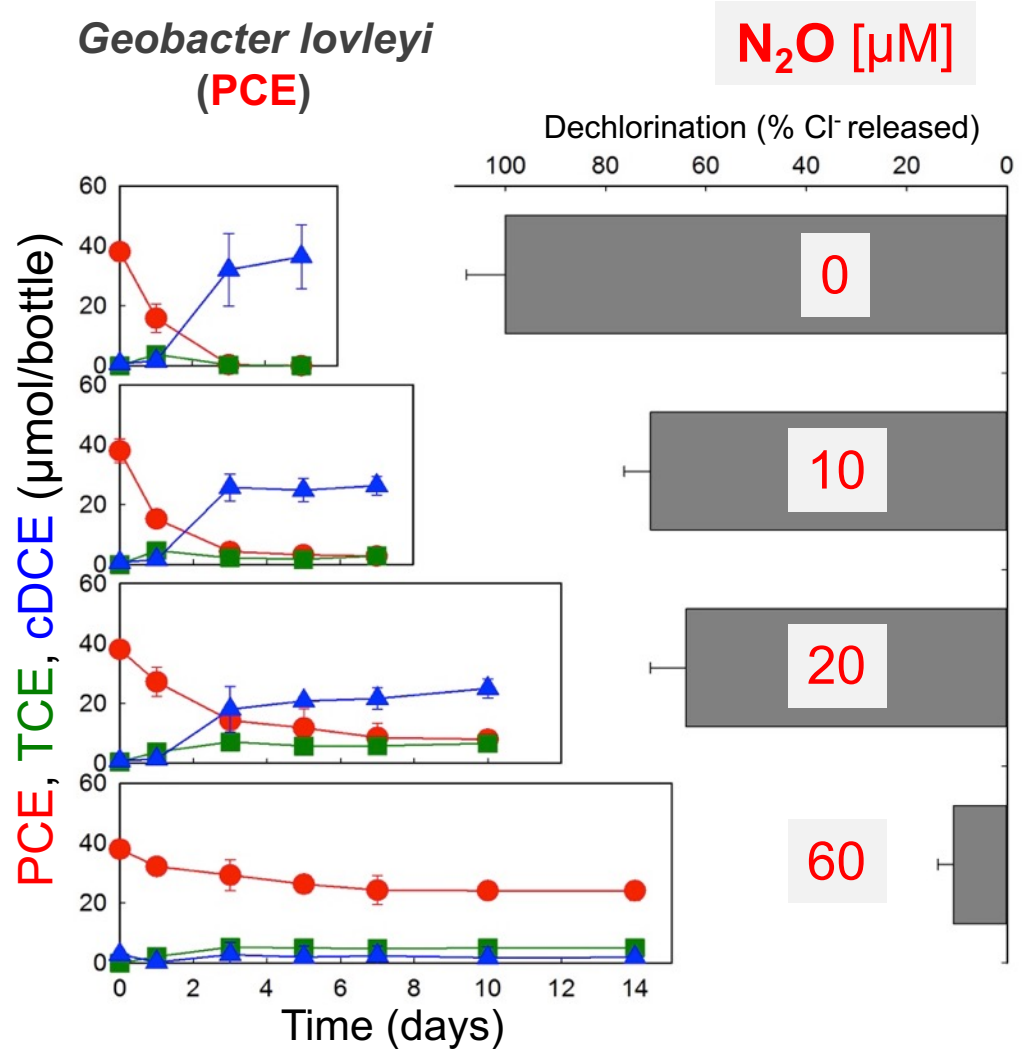
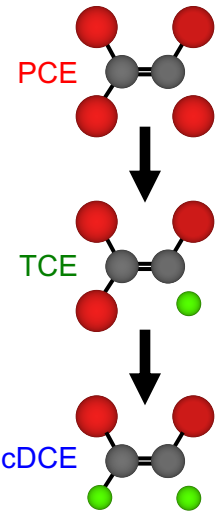
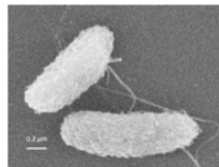


Impact of N₂O on corrinoid-dependent reductive dechlorination?

Jurado et al. 2017, Sci. Total Environ. 584–585:207–218

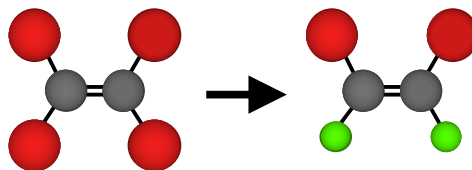


N₂O Inhibits Corrinoid-Dependent Reductive Dechlorination

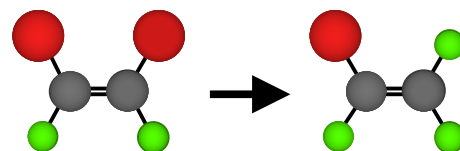


Impact of Increased N₂O in Environmental Systems

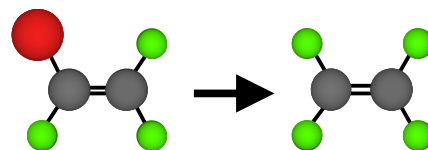
N ₂ O (μM)
0.3
12.5
37.4
75
84
143
65.7



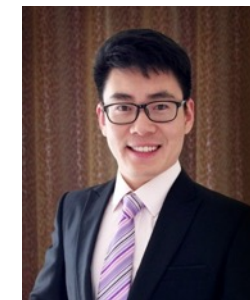
$$K_{i, N_2O} = 40.8 \pm 3.8 \mu\text{M}$$



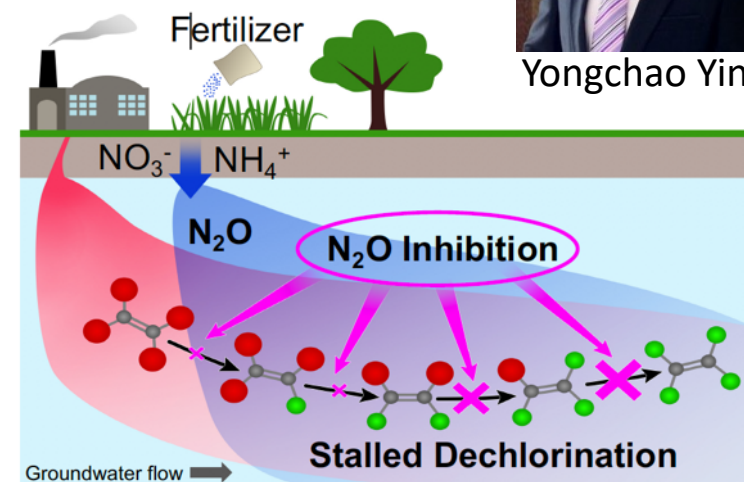
$$K_{i, N_2O} = 21.2 \pm 3.5 \mu\text{M}$$



$$K_{i, N_2O} = 9.6 \pm 0.4 \mu\text{M}$$



Yongchao Yin



Yin et al. 2019. Environ. Sci. Technol. Nitrous Oxide is a Potent Inhibitor of Bacterial Reductive Dechlorination. 53:692-701



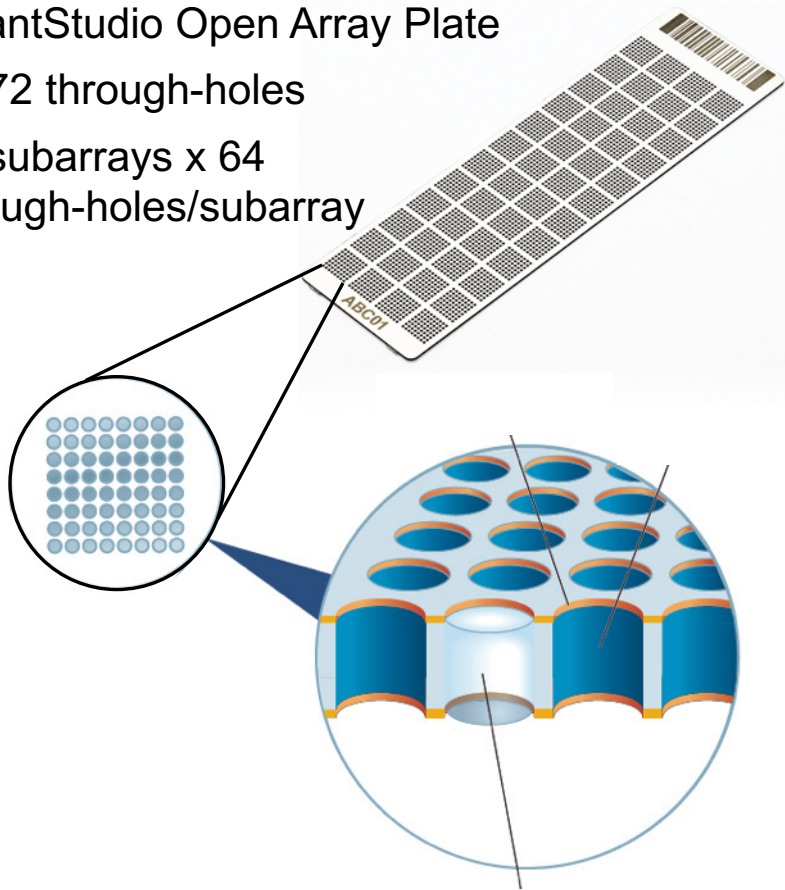
High-Throughput qPCR

- Monitor many biomarker genes simultaneously

QuantStudio Open Array Plate

3,072 through-holes

48 subarrays x 64 through-holes/subarray



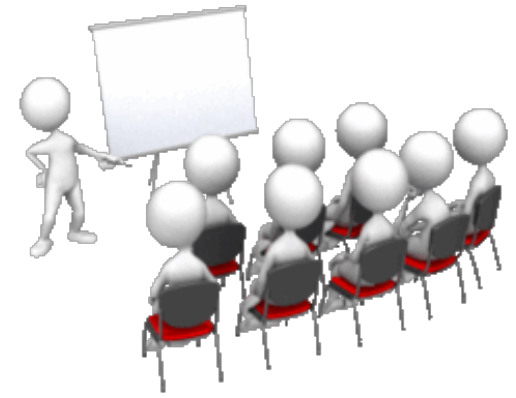
- Scalable platform (224x12; 112x24; 56x48)
- Cost per reaction reduced from \$3.0 to \$0.30
- Four plates can be cycled simultaneously, producing up to 12,288 qPCR data points per run



Robot reduces pipetting errors

Kara Murdoch et al. 2019. In Preparation

Take Home Messages



- Corrinoids are essential for most organisms
- Corrinoid-auxotrophic OHRBs (e.g., *Dhc*, *Dhgm*) are ideal systems to study corrinoid effects on metabolism
- Corrinoid quantity (flux) affects dechlorination activity
- Corrinoid type (lower base) determines reductive dechlorination rates and end points (i.e., function) in *Dhc*
- Geochemistry affects corrinoid pool
- Purine is a naturally occurring lower base
- New avenues to manipulate microbial metabolism (function, ecology → biotechnology, medicine)



Peer-reviewed Manuscripts

Yin, Y., Yan, G. Chen, F. Kara Murdoch, N. Pfisterer, and F.E. Löffler. 2019. Nitrous oxide is a potent inhibitor of bacterial reductive dechlorination. *Environ. Sci. Technol.* 53:692-701 | doi: 10.1021/acs.est.8b05871

Yan, J., M. Bi, A.K. Bourdon, A.T. Farmer, P.-H. Wang, O. Molenda, A. Quaile, N. Jiang, Y. Yang, Y. Yin, B. Şimşir, S.R. Campagna, E.A. Edwards, and F.E. Löffler. 2018. Purinyl-cobamide is a native prosthetic group of reductive dehalogenases. *Nat. Chem. Biol.* 14:8-14. | doi:10.1038/nchembio.2512

Clark, K., D.M. Taggart, B.R. Baldwin, K.M. Ritalahti, R.W. Murdoch, J.K. Hatt, and F.E. Löffler. 2018. Normalized quantitative PCR measurements as predictors for ethene formation at sites impacted with chlorinated ethenes. *Environ. Sci. Technol.* 52:13410-13420 | doi: 10.1021/acs.est.8b04373

Yang, Y., S.A. Higgins, J. Yan, B. Şimşir, K. Chourey, R. Iyer, R.L. Hettich, B. Baldwin, D.M. Ogles, and F.E. Löffler. 2017. Grape pomace compost harbors organohalide-respiring *Dehalogenimonas* species with novel reductive dehalogenase genes. *The ISME Journal.* 11:2767-2780. | doi: 10.1038/ismej.2017.127

Yan, J., B. Şimşir, A.T. Farmer, M. Bi, Y. Yang, S.R. Campagna, and F.E. Löffler. 2016. The corrinoid cofactor of reductive dehalogenases affects dechlorination rates and extents in organohalide-respiring *Dehalococcoides mccartyi*. *ISME J.* 10:1092-1101. | doi: 10.1038/ismej.2015.197



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